

# Main immunoregulatory mechanisms that favor breast cancer development

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## Abstract

Even after the improvements made in recent years in early diagnosis and treatments, breast cancer is still the most common cancer and the leading cause of cancer death in women around the world. Several attempts to design new alternative therapies like immunotherapy have been evaluated in clinical trials, but they have shown limited efficacy. The failure of immunotherapy may be related to suppressive mechanisms in the tumor environment. Consequently, for the development of new immunotherapy based treatment strategies is very important to understand the immunoregulatory mechanisms present in the tumor microenvironment. Some of the immunoregulatory mechanisms described in breast cancer will be discussed in this review.

**KEY WORDS:** Breast cancer. Immunotherapy. Immunoregulation. Immunosuppression.

## Introduction

Cancer is a disease that affects a large number of people and constitutes one of the main causes of death in the entire world. Most important neoplasms include breast cancer, which is the malignancy with the highest incidence and mortality in women worldwide<sup>1</sup>. In Mexico, breast cancer is the malignancy with the highest incidence and mortality<sup>2</sup>. Breast cancer most common type is ductal carcinoma, which originates in cells of the galactophorous ducts that carry milk from the lobules to the nipple. Breast cancer can also originate in cells of the lobules (lobular carcinoma), which are the milk-producing glands. Less often, breast cancer can originate in stromal tissues, including fatty and fibrous connective tissues of the breast<sup>3</sup>. Inflammatory carcinoma is an infrequent breast cancer variant that is highly aggressive, has a 5-year survival rate of 40% and affects especially young women<sup>4,5</sup>. Cancer cells

can invade surrounding healthy breast tissue and reach axillary lymph nodes, thus obtaining a door for access to other parts of the body. In addition to histological classification, there are breast cancer subtypes depending on genetic alterations, gene differential expression and the type of leukocytic infiltrate, among other factors<sup>6</sup>. Together, these factors determine each tumor's biological behavior and treatment response<sup>7</sup>.

Breast cancer treatment depends on the type of cancer and the degree of dissemination<sup>8</sup>. Current treatments are highly aggressive for patients and consist in surgical removal of the tumor and adjuvant therapies such as radiotherapy, chemotherapy, immunotherapy and hormone therapy. Their efficacy largely depends on an early diagnosis enabling small tumors to be eliminated by surgical or chemical methods; however, even at early stages, treatment cannot warrant tumor cell complete elimination, which results in a high rate of disease recurrence. The risk of breast cancer relapse depends both on tumor extent and biological

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characteristics. Lymph node involvement status is one of the most widely used prognostic factors, with a clear correlation existing between the number of lymph nodes involved and the risk for relapse. Recurrent cancer can appear at the same site (local recurrence), in a nearby area such as the chest wall and infraclavicular or supraclavicular lymph nodes (regional recurrence) or at distant sites such as the liver, the lung or the brain (distant recurrence)<sup>9</sup>. Unfortunately, recurrent cancer is highly aggressive and develops rapidly, which largely limits treatment options and efficacy. One treatment alternative that has been investigated in recent years is immunotherapy, but immune tolerance mechanisms that normally protect against the development of autoimmune diseases have been observed to be able to be used by several tumors to evade or suppress local immune response, thereby hindering the development of effective anti-tumor immunity and limiting the therapeutic effects of different immunologic strategies such as the use of dendritic cell-based vaccines<sup>10-12</sup>. Consequently, for the development of new immunotherapy strategies it is highly important to understand the immunoregulatory systems present in tumor microenvironment. Here, some of the immunoregulatory mechanisms described in breast cancer will be discussed.

## **Tumor antigens**

Tumor cells can express proteins that are recognized as antigens by the immune system. These proteins can be tumor-specific antigens (TSA), such as mutated proteins, chromosomal aberrations or some viral proteins' derivatives. There are also the so-called tumor-associated antigens (TAA), which are not tumor-specific and can be found in healthy tissues, but are more commonly expressed by tumor cells than TSA<sup>13</sup>. TAAs may be originated by overexpression of cellular proteins that normally are not exposed to the immune system, such as embryonic proteins<sup>14</sup>. Most TAAs are weak immunogens because they derive from the body's own proteins, which limits immune recognition. In addition, lymphocytes that are able to recognize these TAAs have low affinity owing to thymic tolerance processes<sup>13</sup>. TAAs aforementioned characteristics have hindered the search for therapeutic targets. Nevertheless, some TAAs showing some of the characteristics required to be therapeutically used in breast cancer, such as being immunogenic and being expressed in tumor cells in a significant percentage of patients, have been identified. Among them, one of the most important is human epidermal growth factor receptor 2 (HER2), which is

expressed in approximately 25% of primary tumors. HER2 is a type 1 transmembrane oncoprotein that was initially described in breast cancer pathogenesis in 1987<sup>15</sup>. HER2 expression is associated with poor prognosis, since this kind of tumors are of rapid evolution and dissemination. Currently, monoclonal antibodies against HER2 have been generated, such as trastuzumab, which is a humanized monoclonal antibody that recognizes HER2 extracellular domain and that was approved by the US Food and Drug Administration in 1998<sup>16</sup>. Its clinical use has improved the survival of patients with breast cancer at early and advanced stages<sup>17</sup>. Trastuzumab's mechanism of action consists in preventing HER2 receptor dimerization, thereby blocking cell proliferation-promoting protein kinases signaling and activation. However, nearly 15% of treated patients relapse in a 12-month period<sup>10,18</sup>. The mechanism of resistance to trastuzumab is HER2 alternate dimerization with other proteins of the same family, such as HER3<sup>19</sup>. In view of such treatment resistance, new agents have been developed, with pertuzumab standing out, which is another anti-HER2 humanized monoclonal antibody that recognizes a different epitope than that recognized by trastuzumab. Pertuzumab prevents HER2 dimerization with HER3<sup>20</sup>. Pertuzumab combination with trastuzumab and chemotherapy with docetaxel has demonstrated higher efficacy in the treatment of breast cancer than either agent separately with docetaxel<sup>18,21</sup>. In addition, other anti-HER2 compounds have been developed, such as lapatinib and the trastuzumab-emtansine conjugate, which are mainly used as second-line treatment in case of resistance to trastuzumab in combination with docetaxel<sup>22,23</sup>. In addition to blocking HER2 dimerization, these antibodies activate antibody-dependent cytotoxicity<sup>24</sup>. Therapeutic success of these monoclonal antibodies is also due to the fact that patients with HER2-overexpressing tumors don't possess multiple immunosuppression mechanisms, as it has been observed in patients with HER2-negative tumors<sup>25,26</sup>. Ertumaxomab is a bivalent antibody that recognizes HER2 and CD3, which enables tumor-infiltrating lymphocytes activation, by means of which it might have higher therapeutic effect; currently, it is being assessed to find the maximum dose and clinical efficacy (ClinicalTrials.gov, NCT01569412)<sup>26,27</sup>.

In conclusion, the use of TAA-targeted monoclonal antibodies is a highly promising therapeutic option, but it depends on specific expression of these antigens in tumor cells, as well as on the absence of immunosuppressant mechanisms that might inactivate their antibody-dependent cytotoxicity effect.

## T cell-mediated immunoregulatory mechanisms

Main immunoregulatory mechanisms that have been described in breast cancer include those mediated by T cells. The presence of CD4+ T cells in tumor infiltrate has been associated with negative prognosis, unlike the presence of CD8+ T cells<sup>28</sup>. One of CD4 T cell populations that plays an important role in breast cancer pathogenesis is interleukin 4 and 13 (IL-4 and IL-13)-producing Th2 cells, which promote tumor development by inducing the production of growth factors that accelerate cancer cell proliferation<sup>29-31</sup>. In addition, both these cytokines have been observed to be involved in anti-tumor immune response local suppression<sup>32,33</sup>, in the promotion of metastasis<sup>34</sup> and in tumor cells resistance to apoptosis<sup>35</sup>. IL-13 can be observed both in tumor and tumor adjacent tissues; however, there is higher expression within tumor tissue, which positively correlates with tumor size<sup>36</sup>. IL-13 tumor development-promoting function in breast cancer has been confirmed in animal models, where its biological activity is blocked<sup>29,33</sup>. The presence of the CCL5 chemokine has been described to promote breast cancer growth and metastasis by attracting CD4 lymphocytes, which express the CCR3 receptor, and by promoting their differentiation into Th2 cells<sup>37-40</sup>. By means of T cell response modulation in animal models, decreasing Th2 cell induction and favoring the presence of Th1 cells has been observed to produce an induction of CD8 cytotoxic cells able to control tumor growth<sup>41</sup>. This strategy is being investigated as a treatment alternative.

Other CD4 T cell population that has been associated with tumor development is regulatory T (Treg) cells, characterized by Forkhead Box P3 (FoxP3) transcription factor expression<sup>42</sup>. Treg cells usually participate in peripheral tolerance mechanisms by preventing auto-immune or immune hyperreactivity processes<sup>43</sup>. An important infiltration of Treg FoxP3+ cells within tumors and surrounding regions has been observed in different neoplasms, including breast cancer; specially, when it occurs in tumor-surrounding areas, this infiltration is associated with poor prognosis<sup>44-48</sup>. Breast tumor-derived Treg cells show specificity for TAA and recognize the same epitopes than effector T cells<sup>49</sup>, which evidences their potential participation in anti-tumor response suppression. In support of the above, the ability of tumor Treg cells to suppress the cytotoxic activity of CD8+ T cells that are specific to TAAs such as HER2 has been demonstrated in animal models<sup>50</sup>. Treg

frequency increase does not only occur at the lesion site; higher frequency of these cells in peripheral blood has also been reported in patients with breast cancer, especially at advanced stages<sup>51</sup>. However, in spite of the existence of an increase of Treg cells in breast cancer patients' blood, their activation status and suppressor capacity are not elevated, unlike what occurs in tumor tissue-derived cells, which show highly elevated activation status and suppressor capacity<sup>47,52</sup>. Treg cells are attracted to tumor cell-produced CCL-22 chemokine, which is recognized by Treg cell-expressed CCR4 receptor<sup>32</sup>. In addition to their migration, Treg cell accumulation in tumor microenvironment is due to in situ proliferation of these cells<sup>52</sup>. Molecules with regulatory function that are highly expressed in tumor Treg cells include cytotoxic T lymphocyte-associated antigen 4 (CTLA-4), inducible T-cell co-stimulator (ICOS) and glucocorticoid-induced tumor necrosis factor receptor (GITR)<sup>52-54</sup>. Since these molecules are important to Treg cells functionality and are highly expressed, they have been studied as potential therapeutic targets that allow for tumor Treg cells suppressor effect to be blocked and this way enable the development of an anti-tumor response<sup>55-57</sup>. Currently, an anti-CTLA4 antibody (ipilimumab) is used in clinical practice for the treatment of melanoma, and its effect on breast cancer is being assessed (ClinicalTrials.gov, NCT01502592). In addition, another anti-CTLA-4 antibody, tremelimumab<sup>58</sup>, is being investigated in clinical trials (ClinicalTrials.gov, NCT02536794). Similarly, there are preclinical studies showing that GITR modulation may favor the development of anti-tumor responses<sup>56,59,61</sup>. The combined use of these agents might have an impact on tumor development by inhibiting one of the main suppressor mechanisms in human tumors, such as Treg cells.

## Antigen-presenting cells in breast cancer

Th2 cell generation in breast cancer is induced by dendritic cells (DC) that have been conditioned by the tumor microenvironment and are characterized for expressing OX40L, which is one of the molecules directly implied in CD4+ T cells polarization towards a Th2 profile<sup>62,63</sup>. DC are conditioned in the tumor microenvironment by the presence of thymic stromal lymphoietin (TSLP), which is produced and secreted by tumor cells and makes for DC to express large amounts of OX40L and not to express IL-12<sup>63</sup>. When interacting with CD4+ T cells, tumor DC induce T cells polarization to produce IL-4, IL-13 and tumor necrosis factor alpha (TNF- $\alpha$ ), but

not IL-10<sup>63</sup>; this Th2 cell variant is known as pro-inflammatory Th2 cells<sup>64</sup>. Furthermore, TSLP-exposed DC have been shown to acquire the ability to induce Treg cells from CD4+ T cells<sup>65</sup>. In addition to Th2 and Treg induction, which favor tumor development, breast tumor-isolated DC have their antigenic, phagocytic and maturation capacity decreased, as well as the expression of co-stimulation molecules, such as CD40 and the B7 family<sup>66,67</sup>. On the other hand, plasmacytoid DC (pDC) that infiltrate breast tumors have been observed to have deficient IFN $\alpha$  production caused by higher amounts of transforming growth factor beta (TGF- $\beta$ ) and TNF- $\alpha$  present in tumor microenvironment<sup>53,68</sup>. These cells have elevated capacity to induce and activate Treg lymphocytes, thus eliciting their accumulation and activation at the lesion site<sup>69</sup>. In fact, infiltration of both cell populations in breast tumors has been associated with poor disease prognosis<sup>69-71</sup>. Tumor pDC express elevated levels of ICOS ligand, and it is through this molecule that pDC activate Treg cells, thus causing for them to produce large amounts of IL-10, which has an immunosuppressant effect<sup>69,70</sup>.

Breast tumors cell infiltrate also contains macrophages; these cells are known as tumor-associated macrophages (TAM)<sup>72,73</sup>. The presence of TAM in breast cancer is associated with poor prognosis and higher risk for the development of metastasis<sup>74-76</sup>. Most TAM have an anti-inflammatory M2 phenotype and express elevated levels of the cyclooxygenase 2 (COX-2) enzyme, which leads to the production of prostaglandins that, in turn, favor the generation of Th2 cells and tumor growth<sup>77-79</sup>. TAM have been described to favor tumor cell proliferation and survival by inducing the expression of molecules such as bcl-2<sup>79</sup>. TAM can induce COX-2 expression in tumor cells by means of the production of IL-1 $\beta$ , which induces TAM polarization in M2, thus generating a retro-stimulation system that favors tumor development<sup>78,80</sup>. TAM can directly affect lymphocyte activation by secreting large amounts of IL-10 and TGF- $\beta$ <sup>81</sup>, and promote tumor growth by means of the secretion of growth factors such as vascular endothelial growth factor and epidermal growth factor<sup>82</sup>. They are also able to produce chemokine CCL22 to attract Treg cells into the tumor microenvironment<sup>83</sup>.

### **Immune system cells and tumor stroma cells interactions**

Tumor microenvironment consists of cancer cells, inflammatory cells and stromal cells. The dynamical interactions of cells that form part of tumor microenvironment

determine the environmental conditions under which tumor development is produced<sup>84</sup>. Recent experimental evidence has demonstrated that an important component of tumor stroma is mesenchymal stem cells (MSCs)<sup>85</sup>. MSCs, which originally were reported in bone marrow stroma, have strong attraction for tumor microenvironment and can differentiate into cells that favor the tumor niche<sup>86-89</sup>. These cells are able to promote tumor growth in breast and colon cancer animal models<sup>90-93</sup>. However, the mechanisms whereby they induce tumor development are not known, and it has been suggested that it can be partly mediated by their angiogenic and immunoregulatory properties, which have been described in vitro for bone marrow-derived MSCs<sup>91,94,95</sup>. One of the mechanisms is tumor cell proliferation and migration induction by means of IL-6 and IL-8 secretion<sup>96,97</sup>. Exposure to TNF- $\alpha$ , which is present in tumor microenvironment, has also been reported to induce chemokines CXCL9, CXCL10 and CXCL11 in MSCs, which promotes attraction and mobilization of tumor cells expressing the CXCR3 receptor for these chemokines<sup>98</sup>. Finally, MSCs have been observed to produce large amounts of TGF- $\beta$ , which directly affects leukocyte functionality and induces Treg cells generation in the tumor microenvironment<sup>99</sup>.

### **Blockade of immune checkpoints**

A series of molecules able to suppress local immune response, known as immune checkpoints, are expressed in tumor infiltrate cells<sup>11</sup>. Among the most widely studied, CTLA-4 and programmed cell death 1 (PD-1) are found mainly in patients with melanoma or renal carcinoma<sup>100-102</sup>. For CTLA-4, there are humanized monoclonal antibodies, developed for the treatment of patients with advanced melanoma<sup>100</sup>. In breast cancer, two anti-CTLA-4 antibodies are currently on evaluation process, as previously mentioned. CTLA-4 block inhibits Treg cells suppressor effect, as previously discussed, but it also favors a prolonged activation of effector T cells by hindering CTLA-4-mediated negative regulation, which is expressed in activated T cells. In these, CTLA-4 uses two inhibitory mechanisms: one of them is the transmission of a negative signal through its extracellular region, and the second is competitive antagonism of the co-stimulating signal that is mediated by CD28 and competes for CD80 and CD86 during interaction with antigen-presenting cells<sup>103-106</sup>. PD-1 is an inhibitory receptor expressed in T cells that limits their capacity of response<sup>107</sup>; its ligand PD-L1 is

expressed in tumor cells and tumor lymphocytic infiltrate, and its expression is correlated with poor prognosis in some tumors<sup>12</sup>. In breast cancer, PD-L1 expression in tumor cells is associated with negative clinical and pathological characteristics<sup>108-110</sup>, with metastasis<sup>111</sup> and with decreased frequency of CD8 T cells<sup>112,113</sup>. PD-L1 expression has also been observed in leukocyte infiltrate, but its clinical meaning is controversial<sup>114-116</sup>. Currently, different clinical trials are ongoing with anti-PD-1 antibodies in patients with breast cancer in order to know their safety and clinical efficacy<sup>117</sup>.

### Impact of chemotherapy on anti-tumor immune response

Many of the medications used in chemotherapy have been described to be able to promote anti-tumor immunity mainly due to their capability to induce tumor cell immunogenic death<sup>118,119</sup>. An example is anthracyclines, which cause for tumor cells to expose calreticulin on their surface, as well the release of chromatin-binding high-mobility group B1 protein (HMGB1) and adenosine triphosphate, which together induce DC activation<sup>120-122</sup>. In addition, doxorubicin has been observed to increase the proliferation of CD8 T cells specific for tumor antigens<sup>123</sup>. 5-fluorouracyl stimulates antigen capture and cross presentation in DC<sup>124</sup>, whereas taxanes increase natural killer cells activity and favor leukocyte infiltration<sup>124,126</sup>. Low-dose cyclophosphamide (50 mg/day) inhibits Treg cells activity by decreasing FoxP3 transcription factor<sup>127</sup>; its use has been assessed in combination with trastuzumab and granulocyte and macrophage-colony stimulating factor, with longer survival being observed in those patients who developed an anti-tumor immune response<sup>18</sup>. Finally, trastuzumab and pertuzumab combination therapy has antibody-dependent cellular cytotoxicity as one of its main mechanisms of action<sup>11,129</sup>.

These data clearly illustrate the importance of immune response in the treatment of breast cancer, demonstrating that therapeutic success of many compounds used in chemotherapy is largely due to their capability to stimulate an anti-tumor immune response in patients.

### T cells adoptive transference

With the use of T cells with specific receptors to some tumor antigens, cytotoxic cells that are able to recognize and react to tumor cells can be generated. At its first stages, this technology used lymphocytes isolated from the same tumor leukocyte infiltrate, but this has the

great limitation that sufficient numbers of specific lymphocytes have to be obtained from clinical samples<sup>130</sup>. Subsequently, the generation of chimeric receptors that contain an antigen recognition site and an intracellular signaling domain were generated in order to guarantee lymphocytic activation when in contact with tumor cells<sup>131,132</sup>. The main limitation is that tumor antigens that are different to own antigens have to be found, since own antigens can elicit own material recognition and generate serious toxicity problems and adverse effects<sup>133</sup>. The use of this technology in breast cancer is beginning to be assessed in some clinical trials (clinicaltrials.gov, NCT01837602 and NCT 02547691).

### Conclusions

Breast cancer remains a public health problem that affects a large number of women. Current treatments, both surgical and chemo- and radiotherapeutic, are highly aggressive for patients, and their efficacy is focused on disease early stages, and considerably decreases at advanced stages. These treatments do not always achieve cancer cells complete elimination and the disease can reappear in short time. In view of this panorama, immunotherapy is highly attractive for the treatment of neoplasms, due to immune response specificity, which can prevent the collateral damages observed with chemotherapy and radiotherapy, which affect both body's tumor cells and normal cells. Another important factor is immune memory, which potentially could decrease recurrence rate. However, to date, many immune treatments have shown limited effect on patient survival. This is largely due to the presence of immunosuppressant mechanisms at the lesion site, such as the presence of Treg and Th2 cells, and the production of suppressor cytokines such as IL-10 and TFG- $\beta$ . Identification and characterization of immunoregulatory mechanisms that affect immune response are essential to the design of new therapeutic strategies that can impact on patient survival. Currently, one of the treatments with the highest efficacy in breast cancer is the use of anti-HER2 monoclonal antibodies (trastuzumab and pertuzumab), which in combination with chemotherapy achieve an important reduction of the disease and can in some cases eliminate the totality of tumor cells. However, it is only applicable to patients with tumors that express the HER2 protein. The efficacy of these monoclonal antibodies is a clear example of the potential of the use of immunotherapy in the treatment of cancer. Currently, several antibodies targeted against immunoregulatory molecules, such as

PD-1 and CTLA-4, are being assessed, with the idea of blocking immunosuppressant mechanisms that inhibit the generation of an anti-tumor response. The combined use of immunoregulatory molecule blockers with immune system activators, such as DC-based vaccines or transference of cytotoxic lymphocytes directed against TAA, is one of the most promising immune strategies in the treatment of breast cancer and other neoplasms. In the next few years, onco-immunology will definitively have a very important boom in the development of new treatments against cancer.

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## References

1. WHO. Global Health Estimates: Deaths by Cause, Age, Sex and Country, 2000-2012. Geneva, World Health Organization, 2014.
2. Ferlay J, Soerjomataram I, Ervik M, et al. GLOBOCAN 2012 v1.0. Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer; 2013. Available from: <http://globocan.iarc.fr>
3. Viale G. The current state of breast cancer classification. *Ann Oncol*. 2012;23(Suppl 10): X207-10.
4. González-Angulo AM, Hennessy BT, Broglio K, et al. Trends for inflammatory breast cancer: is survival improving? *Oncologist*. 2007;12:904-12.
5. Cristofanilli M, Valero V, Buzdar AU, et al. Inflammatory breast cancer (IBC) and patterns of recurrence: understanding the biology of a unique disease. *Cancer*. 2007;110:1436-44.
6. Zardavas D, Irtthum A, Swanton C, Piccart M. Clinical management of breast cancer heterogeneity. *Nat Rev Clin Oncol*. 2015;12:381-94.
7. Perou CM, Sorlie T, Eisen MB, et al. Molecular portraits of human breast tumours. *Nature*. 2000;406:747-52.
8. Santa-María CA, Gradishar WJ. Changing treatment paradigms in metastatic breast cancer: lessons learned. *JAMA Oncol*. 2015;1:528-34.
9. Lasso Varela A, Cobos Campos R, Alia Ramos A. Recurrencias loco-regionales en pacientes con cáncer de mama invasivo que presentan 3 ganglios positivos o menos. ¿Está indicada la radioterapia? *Clinica e Investigación en Ginecología y Obstetricia*. 2012;39:203-9.
10. Spector NL, Blackwell KL. Understanding the mechanisms behind trastuzumab therapy for human epidermal growth factor receptor 2-positive breast cancer. *J Clin Oncol*. 2009;27:5838-47.
11. Ravelli A, Reuben JM, Lanza F, et al. Immune-related strategies driving immunotherapy in breast cancer treatment: a real clinical opportunity. *Expert Rev Anticancer Ther*. 2015;15:689-702.
12. Ernst B, Anderson KS. Immunotherapy for the treatment of breast cancer. *Curr Oncol Rep*. 2015;17:5.
13. Finn OJ. Cancer immunology. *N Engl J Med*. 2008;358:2704-15.
14. Criscitiello C. Tumor-associated antigens in breast cancer. *Breast Care (Basel)*. 2012;7:262-6.
15. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science*. 1987;235:177-82.
16. McKeage K, Perry CM. Trastuzumab: a review of its use in the treatment of metastatic breast cancer overexpressing HER2. *Drugs*. 2002; 62:209-43.
17. Maly JJ, Macrae ER. Pertuzumab in combination with trastuzumab and chemotherapy in the treatment of HER2-positive metastatic breast cancer: safety, efficacy, and progression free survival. *Breast Cancer (Auckl)*. 2014;8:81-8.
18. Nahta R, Hung MC, Esteva FJ. The HER-2-targeting antibodies trastuzumab and pertuzumab synergistically inhibit the survival of breast cancer cells. *Cancer Res*. 2004;64:2343-6.
19. Baselga J, Swain SM. Novel anticancer targets: revisiting ERBB2 and discovering ERBB3. *Nat Rev Cancer*. 2009;9:463-75.
20. Franklin MC, Carey KD, Vajdos FF, Leahy DJ, de Vos AM, Sliwkowski MX. Insights into ErbB signaling from the structure of the ErbB2-pertuzumab complex. *Cancer Cell*. 2004;5:317-28.
21. Gianni L, Pienkowski T, Im YH, et al. Efficacy and safety of neoadjuvant pertuzumab and trastuzumab in women with locally advanced, inflammatory, or early HER2-positive breast cancer (NeoSphere): a randomised multicentre, open-label, phase 2 trial. *Lancet Oncol*. 2012;13:25-32.
22. Verma S, Miles D, Gianni L, et al. Trastuzumab emtansine for HER2-positive advanced breast cancer. *N Engl J Med*. 2012;367:1783-91.
23. Lewis Phillips GD, Li G, Dugger DL, et al. Targeting HER2-positive breast cancer with trastuzumab-DM1, an antibody-cytotoxic drug conjugate. *Cancer Res*. 2008;68:9280-90.
24. Musolino A, Naldi N, Bortesi B, et al. Immunoglobulin G fragment C receptor polymorphisms and clinical efficacy of trastuzumab-based therapy in patients with HER-2/neu-positive metastatic breast cancer. *J Clin Oncol*. 2008;26:1789-96.
25. Muraro E, Martorelli D, Turchet E, et al. A different immunologic profile characterizes patients with HER-2-overexpressing and HER-2-negative locally advanced breast cancer: implications for immune-based therapies. *Breast Cancer Res*. 2011;13:R117.
26. Perez EA, Thompson EA, Ballman KV, et al. Genomic analysis reveals that immune function genes are strongly linked to clinical outcome in the North Central Cancer Treatment Group n9831 Adjuvant Trastuzumab Trial. *J Clin Oncol*. 2015;33:701-8.
27. Kiewe P, Hasmmuller S, Kahlert S, et al. Phase I trial of the trifunctional anti-HER2 x anti-CD3 antibody ertumaxomab in metastatic breast cancer. *Clin Cancer Res*. 2006;12:3085-91.
28. Huang Y, Ma C, Zhang Q, et al. CD4+ and CD8+ T cells have opposing roles in breast cancer progression and outcome. *Oncotarget*. 2015;6:17462-78.
29. Aspod C, Pedroza-González A, Gallegos M, et al. Breast cancer instructs dendritic cells to prime interleukin 13-secreting CD4+ T cells that facilitate tumor development. *J Exp Med*. 2007;204:1037-47.
30. Faghiih Z, Erfani N, Haghshenas MR, Safaei A, Talei AR, Ghaderi A. Immune profiles of CD4+ lymphocyte subsets in breast cancer tumor draining lymph nodes. *Immunol Lett*. 2014;158:57-65.
31. Li B, Li Y, Wang XY, et al. Profile of differentially expressed intratumoral cytokines to predict the immune-polarizing side effects of tamoxifen in breast cancer treatment. *Am J Cancer Res*. 2015;5:726-37.
32. Terabe M, Matsui S, Noben-Trauth N, et al. NKT cell-mediated repression of tumor immunosurveillance by IL-13 and the IL-4R-STAT6 pathway. *Nat Immunol*. 2000;1:515-20.
33. Park JM, Terabe M, Donaldson DD, Forni G, Berzofsky JA. Natural immunosurveillance against spontaneous, autochthonous breast cancers revealed and enhanced by blockade of IL-13-mediated negative regulation. *Cancer Immunol Immunother*. 2008;57:907-12.
34. Faghiih Z, Rezaeifard S, Safaei A, Ghaderi A, Erfani N. IL-17 and IL-4 producing CD8+ T cells in tumor draining lymph nodes of breast cancer patients: positive association with tumor progression. *Iran J Immunol*. 2013;10:193-204.
35. Zhang WJ, Li BH, Yang XZ, et al. IL-4-induced Stat6 activities affect apoptosis and gene expression in breast cancer cells. *Cytokine*. 2008;42:39-47.
36. Srabovici N, Mujagic Z, Mujanovic-Mustedanagic J, Muminovic Z, Softic A, Begic L. Interleukin 13 expression in the primary breast cancer tumour tissue. *Biochem Med (Zagreb)*. 2011;21:131-8.
37. Luboshits G, Shina S, Kaplan O, et al. Elevated expression of the CC chemokine regulated on activation, normal T cell expressed and secreted (RANTES) in advanced breast carcinoma. *Cancer Res*. 1999;59:4681-7.
38. Yaal-Hahoshen N, Shina S, Leider-Trejo L, et al. The chemokine CCL5 as a potential prognostic factor predicting disease progression in stage II breast cancer patients. *Clin Cancer Res*. 2006;12:4474-80.
39. Niwa Y, Akamatsu H, Niwa H, Sumi H, Ozaki Y, Abe A. Correlation of tissue and plasma RANTES levels with disease course in patients with breast or cervical cancer. *Clin Cancer Res*. 2001;7:285-9.
40. Zhang Q, Qin J, Zhong L, et al. CCL5-mediated Th2 immune polarization promotes metastasis in luminal breast cancer. *Cancer Res*. 2015;75:4312-21.
41. Wu TC, Xu K, Bancheau R, et al. Reprogramming tumor-infiltrating dendritic cells for CD103+ CD8+ mucosal T-cell differentiation and breast cancer rejection. *Cancer Immunol Res*. 2014;2:487-500.
42. Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science*. 2003;299:1057-61.
43. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell*. 2008;133:775-87.
44. Kim S, Lee A, Lim W, et al. Zonal difference and prognostic significance of foxp3 regulatory T cell infiltration in breast cancer. *J Breast Cancer*. 2014;17:8-17.
45. Takenaka M, Seki N, Toh U, et al. FOXP3 expression in tumor cells and tumor-infiltrating lymphocytes is associated with breast cancer prognosis. *Mol Clin Oncol*. 2013;1:625-32.
46. Lee S, Cho EY, Park YH, Ahn JS, Im YH. Prognostic impact of FOXP3 expression in triple-negative breast cancer. *Acta Oncol*. 2013;52:73-81.

47. Pedroza-González A, Verhoef C, Ijzermans JN, et al. Activated tumor-infiltrating CD4+ regulatory T cells restrain antitumor immunity in patients with primary or metastatic liver cancer. *Hepatology*. 2013;57:183-94.
48. Tang Y, Xu X, Guo S, et al. An increased abundance of tumor-infiltrating regulatory T cells is correlated with the progression and prognosis of pancreatic ductal adenocarcinoma. *PLoS One*. 2014;9:e91551.
49. Schmidt HH, Ge Y, Hartmann FJ, et al. HLA class II tetramers reveal tissue-specific regulatory T cells that suppress T-cell responses in breast carcinoma patients. *Oncoimmunology*. 2013;2:e24962.
50. Weiss VL, Lee TH, Song H, et al. Trafficking of high avidity HER-2/neu-specific T cells into HER-2/neu-expressing tumors after depletion of effector/memory-like regulatory T cells. *PLoS One*. 2012;7:e31962.
51. Wang ZK, Yang B, Liu H, et al. Regulatory T cells increase in breast cancer and in stage IV breast cancer. *Cancer Immunol Immunother*. 2012;61:911-6.
52. Gobert M, Treilleux I, Bendriss-Vermare N, et al. Regulatory T cells recruited through CCL22/CCR4 are selectively activated in lymphoid infiltrates surrounding primary breast tumors and lead to an adverse clinical outcome. *Cancer Res*. 2009;69:2000-9.
53. Sisirak V, Faget J, Gobert M, et al. Impaired IFN-alpha production by plasmacytoid dendritic cells favors regulatory T-cell expansion that may contribute to breast cancer progression. *Cancer Res*. 2012;72:5188-97.
54. Krausz LT, Fischer-Fodor E, Major ZZ, Fetica B. G1TR-expressing regulatory T-cell subsets are increased in tumor-positive lymph nodes from advanced breast cancer patients as compared to tumor-negative lymph nodes. *Int J Immunopathol Pharmacol*. 2012;25:59-66.
55. Le Mercier I, Pujol D, Sanlaville A, et al. Tumor promotion by intratumoral plasmacytoid dendritic cells is reversed by TLR7 ligand treatment. *Cancer Res*. 2013;73:4629-40.
56. Pedroza-González A, Kwekkeboom J, Sprengers D. T-cell suppression mediated by regulatory T cells infiltrating hepatic tumors can be overcome by G1TRL treatment. *Oncoimmunology*. 2013;2:e22450.
57. Pedroza-González A, Zhou G, Singh SP, et al. G1TR engagement in combination with CTLA-4 blockade completely abrogates immunosuppression mediated by human liver tumor-derived regulatory T cells *in vivo*. *Oncoimmunology*. 2015;4:e1051297.
58. Vonderheide RH, LoRusso PM, Khalil M, et al. Tremelimumab in combination with exemestane in patients with advanced breast cancer and treatment-associated modulation of inducible costimulator expression on patient T cells. *Clin Cancer Res*. 2010;16:3485-94.
59. Lu L, Xu X, Zhang B, Zhang R, Ji H, Wang X. Combined PD-1 blockade and G1TR triggering induce a potent antitumor immunity in murine cancer models and synergizes with chemotherapeutic drugs. *J Transl Med*. 2014;12:36.
60. Pruitt SK, Boczkowski D, De Rosa N, et al. Enhancement of anti-tumor immunity through local modulation of CTLA-4 and G1TR by dendritic cells. *Eur J Immunol*. 2011;41:3553-63.
61. Liu Z, Tian S, Falo LD Jr, Sakaguchi S, You Z. Therapeutic immunity by adoptive tumor-primed CD4(+) T-cell transfer in combination with *in vivo* G1TR ligation. *Mol Ther*. 2009;17:1274-81.
62. Liu YJ, Soumelis V, Watanabe N, et al. TSLP: an epithelial cell cytokine that regulates T cell differentiation by conditioning dendritic cell maturation. *Annu Rev Immunol*. 2007;25:193-219.
63. Pedroza-González A, Xu K, Wu TC, et al. Thymic stromal lymphopoietin fosters human breast tumor growth by promoting type 2 inflammation. *J Exp Med*. 2011;208:479-90.
64. Soumelis V, Reche PA, Kanzler H, et al. Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. *Nat Immunol*. 2002;3:673-80.
65. Hanabuchi S, Ito T, Park WR, et al. Thymic stromal lymphopoietin-activated plasmacytoid dendritic cells induce the generation of FOXP3+ regulatory T cells in human thymus. *J Immunol*. 2010;184:2999-3007.
66. Pockaj BA, Basu GD, Pathangey LB, et al. Reduced T-cell and dendritic cell function is related to cyclooxygenase-2 overexpression and prostaglandin E2 secretion in patients with breast cancer. *Ann Surg Oncol*. 2004;11:328-39.
67. Gabrilovich DI, Corak J, Ciernik IF, Kavanaugh D, Carbone DP. Decreased antigen presentation by dendritic cells in patients with breast cancer. *Clin Cancer Res*. 1997;3:483-90.
68. Sisirak V, Faget J, Vey N, et al. Plasmacytoid dendritic cells deficient in IFNalpha production promote the amplification of FOXP3 regulatory T cells and are associated with poor prognosis in breast cancer patients. *Oncoimmunology*. 2013;2:e22338.
69. Faget J, Bendriss-Vermare N, Gobert M, et al. ICOS-ligand expression on plasmacytoid dendritic cells supports breast cancer progression by promoting the accumulation of immunosuppressive CD4+ T cells. *Cancer Res*. 2012;72:6130-41.
70. Faget J, Sisirak V, Blay JY, Caux C, Bendriss-Vermare N, Menetrier-Caux C. ICOS is associated with poor prognosis in breast cancer as it promotes the amplification of immunosuppressive CD4 T cells by plasmacytoid dendritic cells. *Oncoimmunology*. 2013;2:e23185.
71. Treilleux I, Blay JY, Bendriss-Vermare N, et al. Dendritic cell infiltration and prognosis of early stage breast cancer. *Clin Cancer Res*. 2004;10:7466-74.
72. Komohara Y, Fujiwara Y, Ohnishi K, Takeya M. Tumor-associated macrophages: potential therapeutic targets for anti-cancer therapy. *Adv Drug Deliv Rev*. 2016;99:180-5.
73. Brady NJ, Chuntova P, Schwertfeger KL. Macrophages: regulators of the inflammatory microenvironment during mammary gland development and breast cancer. *Mediators Inflamm*. 2016;2016:4549676.
74. Gwak JM, Jang MH, Kim DI, Seo AN, Park SY. Prognostic value of tumor-associated macrophages according to histologic locations and hormone receptor status in breast cancer. *PLoS One*. 2015;10:e0125728.
75. Yuan ZY, Luo RZ, Peng RJ, Wang SS, Xue C. High infiltration of tumor-associated macrophages in triple-negative breast cancer is associated with a higher risk of distant metastasis. *Onco Targets Ther*. 2014;7:1475-80.
76. Mahmoud SM, Lee AH, Paish EC, Macmillan RD, Ellis IO, Green AR. Tumor-infiltrating macrophages and clinical outcome in breast cancer. *J Clin Pathol*. 2012;65:159-63.
77. Kalinski P, Hilkens CM, Sniijders A, Sniijdewint FG, Kapsenberg ML. IL-12-deficient dendritic cells, generated in the presence of prostaglandin E2, promote type 2 cytokine production in maturing human naive T helper cells. *J Immunol*. 1997;159:28-35.
78. Li H, Yang B, Huang J, et al. Cyclooxygenase-2 in tumor-associated macrophages promotes breast cancer cell survival by triggering a positive-feedback loop between macrophages and cancer cells. *Oncotarget*. 2015;6:29637-50.
79. Sousa S, Brion R, Lintunen M, et al. Human breast cancer cells educate macrophages toward the M2 activation status. *Breast Cancer Res*. 2015;17:101.
80. Hou Z, Falcone DJ, Subbaramaiah K, Dannenberg AJ. Macrophages induce COX-2 expression in breast cancer cells: role of IL-1beta autoamplification. *Carcinogenesis*. 2011;32:695-702.
81. Chanmee T, Ontong P, Konno K, Itano N. Tumor-associated macrophages as major players in the tumor microenvironment. *Cancers (Basel)*. 2014;6:1670-90.
82. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. *Immunity*. 2014;41:49-61.
83. Ostuni R, Kratochvill F, Murray PJ, Natoli G. Macrophages and cancer: from mechanisms to therapeutic implications. *Trends Immunol*. 2015;36:229-39.
84. Protti MP, De Monte L. Cross-talk within the tumor microenvironment mediates Th2-type inflammation in pancreatic cancer. *Oncoimmunology*. 2012;1:89-91.
85. Hall B, Andreeff M, Marini F. The participation of mesenchymal stem cells in tumor stroma formation and their application as targeted-gene delivery vehicles. *Handb Exp Pharmacol*. 2007;(180):263-83.
86. Direkze NC, Hoidalva-Dilke K, Jeffery R, et al. Bone marrow contribution to tumor-associated myofibroblasts and fibroblasts. *Cancer Res*. 2004;64:8492-5.
87. Kidd S, Spaeth E, Watson K, et al. Origins of the tumor microenvironment: quantitative assessment of adipose-derived and bone marrow-derived stroma. *PLoS One*. 2012;7:e30563.
88. Mishra PJ, Mishra PJ, Humeniuk R, et al. Carcinoma-associated fibroblast-like differentiation of human mesenchymal stem cells. *Cancer Res*. 2008;68:4331-9.
89. Quante M, Tu SP, Tomita H, et al. Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. *Cancer Cell*. 2011;19:257-72.
90. Dvorak HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med*. 1986;315:1650-9.
91. Hogan NM, Dwyer RM, Joyce MR, Kerin MJ. Mesenchymal stem cells in the colorectal tumor microenvironment: recent progress and implications. *Int J Cancer*. 2012;131:1-7.
92. Karnoub AE, Dash AB, Vo AP, et al. Mesenchymal stem cells within tumor stroma promote breast cancer metastasis. *Nature*. 2007;449:557-63.
93. Shinagawa K, Kitadai Y, Tanaka M, et al. Mesenchymal stem cells enhance growth and metastasis of colon cancer. *Int J Cancer*. 2010;127:2323-33.
94. Peddaredidigari VG, Wang D, Dubois RN. The tumor microenvironment in colorectal carcinogenesis. *Cancer Microenviron*. 2010;3:149-66.
95. Hernanda PY, Pedroza-González A, van der Laan LJ, et al. Tumor promotion through the mesenchymal stem cell compartment in human hepatocellular carcinoma. *Carcinogenesis*. 2013;34:2330-40.
96. Ma F, Chen D, Chen F, et al. Human umbilical cord mesenchymal stem cells promote breast cancer metastasis by interleukin-8 and interleukin-6 dependent induction of CD44/CD24 cells. *Cell Transplant*. 2015;24:2585-99.
97. Di GH, Liu Y, Lu Y, Liu J, Wu C, Duan HF. IL-6 secreted from senescent mesenchymal stem cells promotes proliferation and migration of breast cancer cells. *PLoS One*. 2014;9:e113572.
98. Shin SY, Nam JS, Lim Y, Lee YH. TNFalpha-exposed bone marrow-derived mesenchymal stem cells promote locomotion of MDA-MB-231 breast cancer cells through transcriptional activation of CXCR3 ligand chemokines. *J Biol Chem*. 2010;285:30731-40.

99. Patel SA, Meyer JR, Greco SJ, Corcoran KE, Bryan M, Rameshwar P. Mesenchymal stem cells protect breast cancer cells through regulatory T cells: role of mesenchymal stem cell-derived TGF-beta. *J Immunol.* 2010;184:5885-94.
100. Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med.* 2010;363:711-23.
101. Sznol M, Chen L. Antagonist antibodies to PD-1 and B7-H1 (PD-L1) in the treatment of advanced human cancer. *Clin Cancer Res.* 2013;19:1021-34.
102. Nahas GR, Walker ND, Bryan M, Rameshwar P. A perspective of immunotherapy for breast cancer: lessons learned and forward directions for all cancers. *Breast Cancer (Auckl).* 2015;9(Suppl 2):35-43.
103. Baroja ML, Vijaykrishnan L, Bettelli E, et al. Inhibition of CTLA-4 function by the regulatory subunit of serine/threonine phosphatase 2A. *J Immunol.* 2002;168:5070-8.
104. Tai X, Van Laethem F, Pobezinsky L, et al. Basis of CTLA-4 function in regulatory and conventional CD4(+) T cells. *Blood.* 2012;119:5155-63.
105. Ise W, Kohyama M, Nutsch KM, et al. CTLA-4 suppresses the pathogenicity of self antigen-specific T cells by cell-intrinsic and cell-extrinsic mechanisms. *Nat Immunol.* 2010;11:129-35.
106. Schneider H, Rudd CE. Diverse mechanisms regulate the surface expression of immunotherapeutic target CTLA-4. *Front Immunol.* 2014;5:619.
107. Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. *EMBO J.* 1992;11:3887-95.
108. Cimino-Mathews A, Thompson E, Taube JM, et al. PD-L1 (B7-H1) expression and the immune tumor microenvironment in primary and metastatic breast carcinomas. *Hum Pathol.* 2016;47:52-63.
109. Baptista MZ, Sarian LO, Derchain SF, Pinto GA, Vassallo J. Prognostic significance of PD-L1 and PD-L2 in breast cancer. *Hum Pathol.* 2016;47:78-84.
110. Mittendorf EA, Philips AV, Meric-Bernstam F, et al. PD-L1 expression in triple-negative breast cancer. *Cancer Immunol Res.* 2014;2:361-70.
111. Mazel M, Jacot W, Pantel K, et al. Frequent expression of PD-L1 on circulating breast cancer cells. *Mol Oncol.* 2015;9:1773-82.
112. Park IH, Kong SY, Ro JY, et al. Prognostic implications of tumor-infiltrating lymphocytes in association with programmed death ligand 1 expression in early-stage breast cancer. *Clin Breast Cancer.* 2016;16:51-8.
113. Schalper KA. PD-L1 expression and tumor-infiltrating lymphocytes: revisiting the antitumor immune response potential in breast cancer. *Oncoimmunology.* 2014;3:e29288.
114. Sabatier R, Finetti P, Mamessier E, et al. Prognostic and predictive value of PDL1 expression in breast cancer. *Oncotarget.* 2015;6:5449-64.
115. Muenst S, Schaerli AR, Gao F, et al. Expression of programmed death ligand 1 (PD-L1) is associated with poor prognosis in human breast cancer. *Breast Cancer Res Treat.* 2014;146:15-24.
116. Muenst S, Soysal SD, Gao F, Obermann EC, Oertli D, Gillanders WE. The presence of programmed death 1 (PD-1)-positive tumor-infiltrating lymphocytes is associated with poor prognosis in human breast cancer. *Breast Cancer Res Treat.* 2013;139:667-76.
117. Chawla A, Philips AV, Alatrash G, Mittendorf E. Immune checkpoints: a therapeutic target in triple negative breast cancer. *Oncoimmunology.* 2014;3:e28325.
118. Criscitiello C, Curigliano G. Immunotherapeutics for breast cancer. *Curr Opin Oncol.* 2013;25:602-8.
119. Tesniere A, Panaretakis T, Kepp O, et al. Molecular characteristics of immunogenic cancer cell death. *Cell Death Differ.* 2008;15:3-12.
120. Sukkurwala AQ, Adjemian S, Senovilla L, et al. Screening of novel immunogenic cell death inducers within the NCI Mechanistic Diversity Set. *Oncoimmunology.* 2014;3:e28473.
121. Ghiringhelli F, Apetoh L, Tesniere A, et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1beta-dependent adaptive immunity against tumors. *Nat Med.* 2009;15:1170-8.
122. Obeid M, Tesniere A, Ghiringhelli F, et al. Calreticulin exposure dictates the immunogenicity of cancer cell death. *Nat Med.* 2007;13:54-61.
123. Mattarollo SR, Loi S, Duret H, Ma Y, Zitvogel L, Smyth MJ. Pivotal role of innate and adaptive immunity in anthracycline chemotherapy of established tumors. *Cancer Res.* 2011;71:4809-20.
124. Galetto A, Buttiglieri S, Forno S, Moro F, Mussa A, Matera L. Drug- and cell-mediated antitumor cytotoxicities modulate cross-presentation of tumor antigens by myeloid dendritic cells. *Anticancer Drugs.* 2003;14:833-43.
125. Carson WE 3rd, Shapiro CL, Crespin TR, Thornton LM, Andersen BL. Cellular immunity in breast cancer patients completing taxane treatment. *Clin Cancer Res.* 2004;10:3401-9.
126. Demaria S, Volm MD, Shapiro RL, et al. Development of tumor-infiltrating lymphocytes in breast cancer after neoadjuvant paclitaxel chemotherapy. *Clin Cancer Res.* 2001;7:3025-30.
127. Ghiringhelli F, Menard C, Puig PE, et al. Metronomic cyclophosphamide regimen selectively depletes CD4+CD25+ regulatory T cells and restores T and NK effector functions in end stage cancer patients. *Cancer Immunol Immunother.* 2007;56:641-8.
128. Chen G, Gupta R, Petrik S, et al. A feasibility study of cyclophosphamide, trastuzumab, and an allogeneic GM-CSF-secreting breast tumor vaccine for HER2+ metastatic breast cancer. *Cancer Immunol Res.* 2014;2:949-61.
129. Yamashita-Kashima Y, Iijima S, Yorozu K, et al. Pertuzumab in combination with trastuzumab shows significantly enhanced antitumor activity in HER2-positive human gastric cancer xenograft models. *Clin Cancer Res.* 2011;17:5060-70.
130. Kunert A, Straetemans T, Govers C, et al. TCR-engineered T cells meet new challenges to treat solid tumors: choice of antigen, T cell fitness, and sensitization of tumor milieu. *Front Immunol.* 2013;4:363.
131. Grupp SA, Kalos M, Barrett D, et al. Chimeric antigen receptor-modified T cells for acute lymphoid leukemia. *N Engl J Med.* 2013;368:1509-18.
132. Kalos M, Levine BL, Porter DL, et al. T cells with chimeric antigen receptors have potent antitumor effects and can establish memory in patients with advanced leukemia. *Sci Transl Med.* 2011;3:95ra73.
133. Maus MV, Haas AR, Beatty GL, et al. T cells expressing chimeric antigen receptors can cause anaphylaxis in humans. *Cancer Immunol Res.* 2013;1:26-31.